Total Synthesis of Bistratamide G, a Metabolite of the Philippines Ascidian *Lissoclinum bistratum*, from Dehydrotripeptides

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Total synthesis of triheterocyclic cyclopeptide bistratamide G, a metabolite from the Philippines ascidian *Lissoclinum bistratum*, was first achieved from two kinds of dehydrotripeptides.

Bistratamide G (1),^{1,2} one of many bistratamide-type polyheterocyclic cyclopeptides, isolated recently from the southern Philippines ascidian *Lissoclinum bistratum*, shows activity in the human colon tumor HCT-116 cell line assay. The natural **1** features an interesting macrocyclic structure constituted of three kinds of heterocyclic (thiazole and oxazole) amino acid residues, as shown in Figure 1.

So far, various useful synthetic methods for the above-mentioned heterocyclic amino acids, and other similar amino acids have been already reported by a number of workers.³ Accordingly, the total synthesis of bistratamide-type dendroamide A and similar natural products has been also achieved by the stepwise elongation of the above amino acids and then by macrocyclization by four groups of investigators.^{4–7} However, though such syntheses have been already reported, for the interesting bioactivities of the polyheterocyclic cyclopeptides, here, a new and general synthetic method for the bistratamides from Δ^2 -and Δ^3 -dehydrotripeptides⁸ has been developed and achieved. In particular, these elongation reactions were carried out without any special condensing agent except for the last macrocyclization



Figure 1. Retrosynthesis of bistratamide G (1).

The syntheses of two kinds of building blocks, (*S*)-2-[1-(*N*-Cbzamino)-2-methylpropyl]oxazole-L-valine thioamide (**2**) and methyl (*S*)-2-{1-[*N*-(3-bromo-2-oxopropanoyl)amino]-2-methylpropyl}-5-methyloxazole-4-carboxylate (**3**) as the left- and right-half components of the linear triheterocyclic peptide **18** as the precursor of **1**, respectively, were accomplished as follows. First of all, to synthesize **2**, the starting Δ^2 -dehydrotripeptide⁸ (Cbz-L-Val- Δ Ala-L-Val-OMe (**5**), Cbz = benzyloxycarbonyl, Δ Ala = α -dehydroalanine residue) was prepared by the usual dehydration of the Ser residue of Cbz-L-Val-L-Ser-L-Val-OMe (**4**) with methanesulfonyl chloride (MsCl) in the presence of Et₃N and then with DBU (1,8-diazabicyclo[5.4.0]undec-7-

ene).⁹ Subsequently, according to the method reported previously,¹⁰ simultaneous bromination and methoxylation of **5** with NBS (*N*-bromosuccinimide) in MeOH gave the corresponding α -methoxylated β -bromoalanyltripeptide **6**. Oxazolination of **6** with Cs₂CO₃ in dioxane gave the corresponding 4-methoxyoxazoline derivative **7**, the methoxy group of which was eliminated with camphorsulfonic acid (CSA) to give the expected oxazole derivative **8**. After hydrolysis of the methyl ester of **8** with 1 M LiOH, the obtained hydrolysate **9** was amidated with CICOOEt in the presence of Et₃N and then with 28% aq. NH₃ to give the corresponding carboxamide **10**. Finally, thiocarbonylation of **10** with Lawesson's reagent gave the required thiocarboxamide **2**, as shown in Scheme 1.



Scheme 1. Reagents and conditions: i) MsCl, Et_3N , DBU, ii) NBS, MeOH, rt, 1 h, iii) Cs₂CO₃, dioxane, 60 °C, 6 h, iv) CSA, toluene, 70 °C, v) 1M LiOH, vi) ClCOOEt, Et_3N , then 28% aq. NH₃, vii) Lawesson's Reagent.

On the other hand, to synthesize 3, firstly, similarly to the case of 5, N,O-isopropylidene (Ip)-N-t-butoxycarbonyl (Boc)-L-Ser-L-Val-(Z)- Δ Abu-OMe (12) (Δ Abu = 2-amino-2-butenoic acid) was prepared by the dehydration of the Thr residue of N,O-Ip-N-Boc-L-Ser-L-Val-L-Thr-OMe (11).⁹ In the case of the bromination of the \triangle Abu residue, it has been already reported that the direct oxazolation of the Δ Abu residue-containing peptide with NBS and then with Cs₂CO₃ proceeds smoothly.^{9,10} Therefore, the bromination of 12 with NBS in CHCl₃, instead of MeOH, was performed to give Δ^3 -dehydrotripeptide 13⁸ containing β -Br- Δ Abu residue, which was then oxazolated with Cs_2CO_3 to give the desired 5-methyloxazole derivative 14. Secondly, selective β -elimination of the Ip group with trifluoroacetic acid (TFA) and CHCl₃ (4:96 v/v), followed by dehydration of the formed N-Boc-seryldipeptide 15 was carried out, similarly to the cases of 5 and 12. That is, dehydration of the Ser residue of 15 and bromination of the obtained Δ^2 -dehydrodipeptide 16⁸ with NBS in MeOH were carried out consecutively to give the corresponding α -methoxylated β -bromoalanyldipeptide derivative 17. Thirdly, deprotection of the Boc group of 17 with TFA and then hydrolysis proceeded smoothly to give 3, as shown in Scheme 2. However, because of its lability, without purification the formed 3 was used in the next thiazolation with 2.



Scheme 2. Reagents and conditions: i) MsCl, Et₃N, CHCl₃, 0° C, 1 h, then DBU, rt, 30 min, ii) NBS, CHCl₃, rt, 1 h, Et₃N, rt, 30 min, iii) Cs₂CO₃, dioxane, 60 °C, 6 h, iv) TFA:CHCl₃ (4:96 v/v), rt, 5 h, v) NBS, MeOH, rt, 30 min, vi) TFA, rt, 30 min, then H₂O, rt, 10 min.

Finally, thiazolation between the thiocarboxamide group of **2** and the bromoacyl group of **3** by successive treatments with KHCO₃ in dimethoxyethane (DME), with trifluoroacetic anhydride (TFAA) and pyridine, and then with 28% aq. NH₃ gave a linear triheterocyclic peptide **18**.¹¹ Hydrolysis of the ester with 1 M LiOH and deprotection of the Cbz group of the hydrolysate **19** with 10% Pd-C/H₂, followed finally by macrocyclization of the obtained *N*,*O*-deprotected triheterocyclic peptide **20** with BOP and (*i*-Pr)₂NEt in DMF under high-dilution conditions (1 mmol/L) at room temperature for 12 h gave the expected **1**¹² in 51% yield from **20**, as shown in Scheme 3.



Scheme 3. Reagents and conditions: i) KHCO₃, DME, 0 °C, 30 min, 50 °C, overnight, b) TFAA, pyridine, 0 °C, 1 h, c) 28% aq. NH₃, 0 °C, ii) a) 1 M LiOH, H₂O-dioxane (1:1 v/v), rt, 30 min, b) 10% Pd-C, H₂, rt, 3 h, iii) BOP, (*i*-Pr)₂NEt, DMF, 12 h.

The structures of all new products thus obtained were confirmed by the ¹H and ¹³C NMR spectral data and the satisfactory results of the elemental analyses. In particular, the chemical and physical constants of the synthetic **1** ($[\alpha]_D^{26} - 82.3^\circ$ (*c* 1.00, MeOH)) were fully identical with those of the natural **1** ($[\alpha]_D$ -73.8° (*c* 1.0, MeOH)).

In conclusion, a convenient and general synthetic method

for the various bistratamide-type metabolites has been efficiently developed.

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- 11 **18**: Colorless solid. mp 71.0–71.5 °C. $[\alpha]_D^{26}$ +4.2° (*c* 0.93, CHCl₃). ¹H NMR (CDCl₃, 600 MHz) $\delta = 0.91$ (d, 3H, J = 6.6 Hz), 0.93 (d, 3H, J = 6.6 Hz), 0.95 (d, 3H, J = 7.2 Hz), 0.97 (d, 3H, J = 7.2 Hz), 0.99 (d, 3H, J = 7.2 Hz), 1.12 (d, 3H, J = 7.2 Hz), 2.23–2.25, 2.35–2.37 and 2.56–2.62 (each m, 1H × 3), 2.61 (s, 3H), 3.91 (s, 3H), 4.84–4.88 (m, 1H), 5.08–5.14 (m, 2H), 5.17–5.20, 5.32–5.38, and 5.49–5.52 (each m, 1H × 2), T_{33} –7.36 (m, 5H), 7.45 and 7.85 (each br d, 1H × 2, J = 9.6 Hz), 8.03 (s, 1H), 8.17 (s, 1H). Found: C, 58.83; H, 6.23; N, 12.48%. Calcd for C₃₄H₄₂N₆O₈S: C, 58.77; H, 6.09; N, 12.19%.
- 12 1: Colorless solid. mp 85.0–85.5 °C. $[\alpha]_D^{26}$ –82.3° (c 1.00, MeOH). IR (KBr) 3396, 2964, 1685, 1676, 1597, 1533, 1508 cm⁻¹. ¹H NMR (DMSO- d_6 , 600 MHz) $\delta = 0.79$ (d, 3H, J = 6.6 Hz), 0.80 (d, 3H, J = 6.6 Hz), 0.83 (d, 3H, J = 7.2 Hz, 0.84 (d, 3H, J = 7.2 Hz), 0.86 (d, 3H, J =7.2 Hz), 0.87 (d, 3H, J = 7.2 Hz), 2.04–2.09 (m, 1H), 2.11– 2.14 (m, 1H), 2.17-2.21 (m, 1H), 2.46 (s, 3H), 4.91 (dd, 1H, J = 7.2, 4.2 Hz, 4.97 (dd, 1H, J = 9.0, 6.0 Hz), 5.28 (dd, 1H, J = 9.0, 6.0 Hz), 8.20 (br d, 1H, J = 7.2 Hz), 8.22 (br d, 1H, J = 9.0 Hz), 8.23 (s, 1H), 8.36 (br d, 1H, J =9.0 Hz), 8.68 (s, 1H). ¹³C NMR (DMSO- d_6 , 150 MHz) $\delta =$ 11.1, 18.0, 18.1, 18.1, 18.2, 18.3, 18.6, 32.6, 32.9, 34.6, 52.1, 52.7, 54.8, 125.2, 128.0, 134.5, 143.0, 147.9, 152.9, 158.4, 159.3, 160.3, 160.6, 163.2, 168.3. MALDI-TOFMS Found: m/z 528.10 (M + H⁺). Calcd for C₂₅H₃₂N₆O₅S: $528.63 (M + H^+).$